# In Situ Data Analytics and Indexing of Protein Trajectories

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# Acknowledgements

#### **Sponsors:**









#### **Collaborators:**





Travis J. Boy

Boyu Z.

Trilce E.

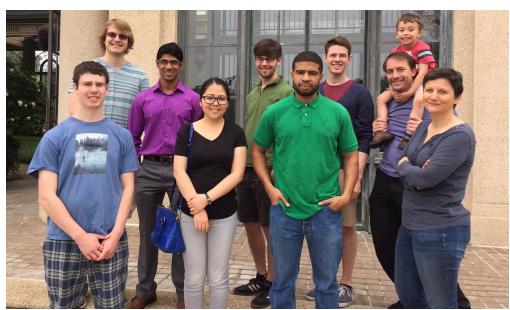




Adam L.

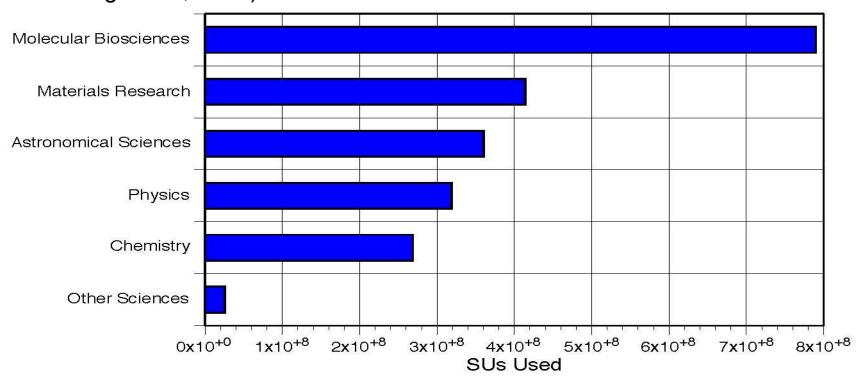
Silvia C.

#### The GCLab@UD



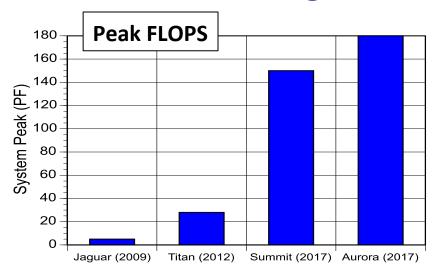
#### MD simulations are alive and kicking!

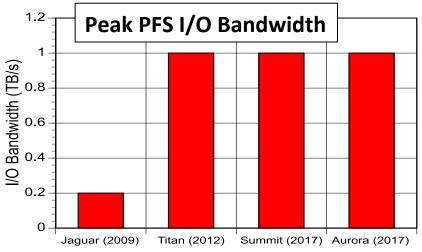
XSEDE SUs used by type of targeted science over the past 6 months (March 1, 2016 - August 31, 2016)



Four of the top 10 XSEDE users run molecular simulations (i.e., Schulten at UIUC, Feig at Michigan State U, Voth at U Chicago, and Case at Rutgers U)

#### Challenges at the Extreme Scale





MD simulations today:

Save all the data to analyze later!

MD simulations at exascale:

- Analyze data as they are generated
- Save only what is really needed!
- Data must be easy to retrieve and reuse

We must change how we run our simulaitons at the exascale

#### **Perspectives**

#### The scientist:

"Storage technologies are advancing [...] and it is really not clear at all [to me] that especially distributed storage platforms would not be able to handle [...] petabyte data sets"

Anonymous Feedback

The computer architect:

"[...] there will be burst buffers on the DOE machines which will give applications much faster I/O [...]"

Anonymous Feedback

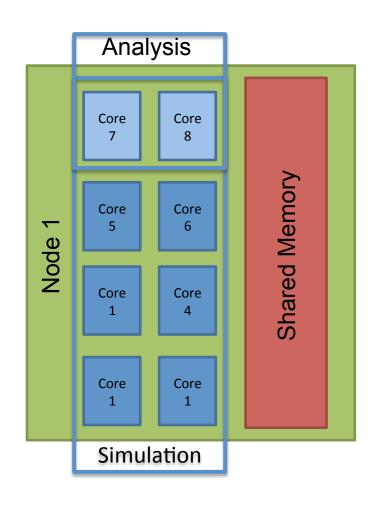
#### **Burst Buffers**

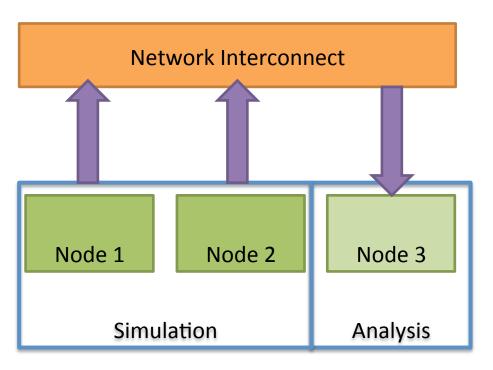
Many have heard about it, few have seen real machines with it, even fewer have ran applications on those machines ...

#### Remaining Challenges

- Burst Buffers are NOT the magic I/O silver bullet
  - I/O contention still a problem if we exceed the burst buffer capability
  - Burst buffers improve offloading bandwidth but do NOT help uploading data from storage for analysis and visualization

#### In-situ and In-transit Analysis





#### Example of tools:

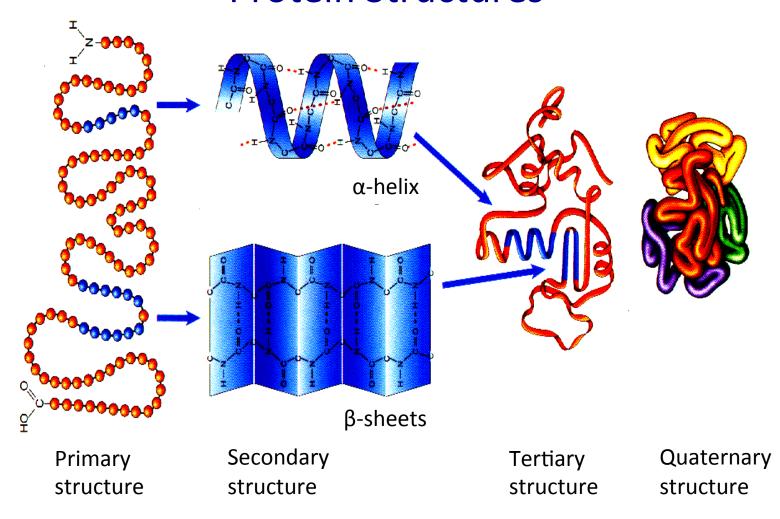
- DataSpaces (Rutgers U.)
- DataStager (GeorgiaTech)

#### In-situ and In-transit MD Analysis

"In-Situ Data Analysis and Indexing of Protein Trajectories," Travis Johnston, Buyu Zhang, Adam Liwo, Silvia Crivelli, and Michela Taufer. JCC 2017.

- Integrate in-situ and in-transit analysis in MD simulations
  - Mitigate data movements by moving the analysis close to the simulation
- Define in-situ algorithms for runtime analysis of molecular structures in molecular dynamics simulations
  - Capture conformation changes of individual secondary structures
  - Capture changes of tertiary structures (i.e., a secondary structure with respect to other secondary structures)

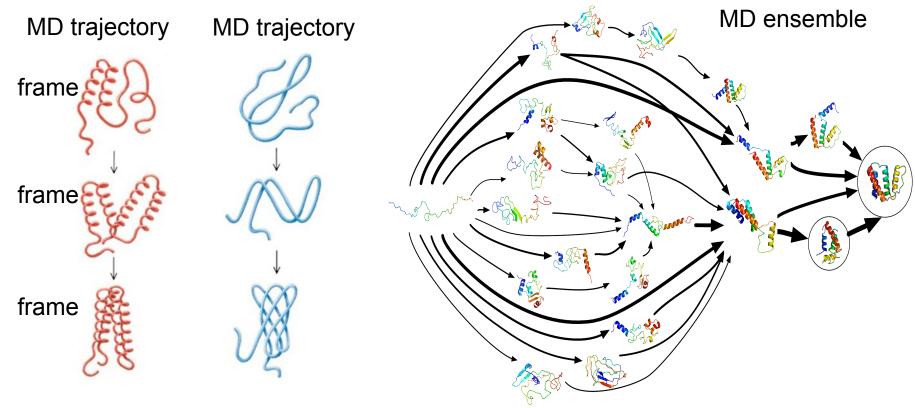
#### **Protein Structures**



From: http://schoolworkhelper.net/protein-structures-primary-secondary-tertiary-quaternary/

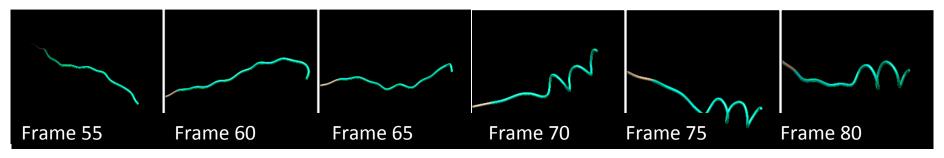
#### **Molecular Dynamics Simulations**

Use computer simulations to study the physical movements of atoms and molecules



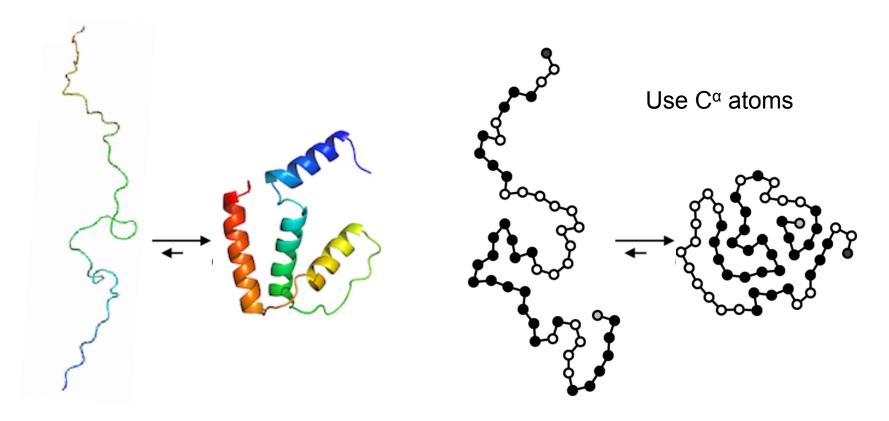
#### **Analysis Requirements**

Frames of an MD trajectory:



- We want to capture what is going on in each frame without:
  - Disrupting the simulation (e.g., stealing CPU and memory on the node)
  - Moving all the frames to a central file system and analyzing them once the simulation is over
  - Comparing each frame with past frames of the same job
  - Comparing each frame with frames of other jobs

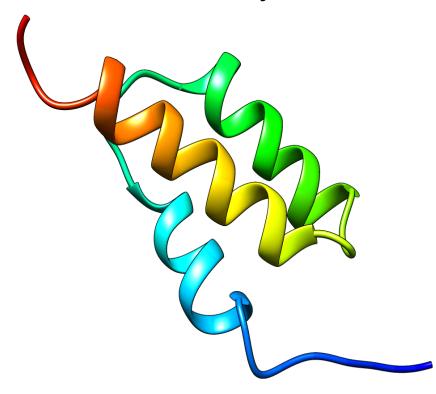
### **Modeling Molecules**



From: By Tomixdf (talk) - Own work (Original text: self-made), Public Domain, https://commons.wikimedia.org/w/index.php?curid=23662306

#### **Capturing Secondary Structures**

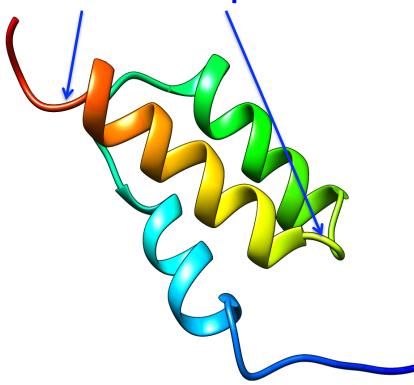
Given a frame of an MD job at time t



#### **Capturing Secondary Structures**

Define the substructure:

start and stop amino acids

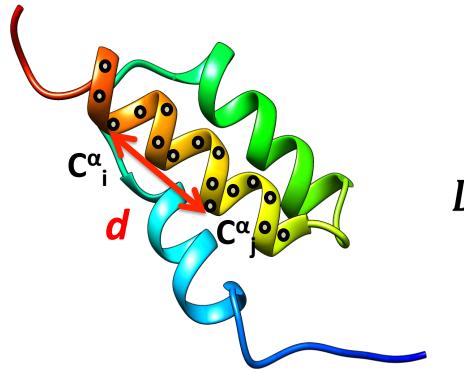


#### **Capturing Secondary Structures**

Measure the distance between  $C^{\alpha}_{i}$  and  $C^{\alpha}_{i}$ 

Build the substructure

Euclidean Distance Matrix (D)



$$D = \begin{bmatrix} 0 & \times & \times & \times & \times & \times \\ \times & 0 & d & \times & \times & \times \\ \times & d & 0 & \times & \times & \times \\ \times & \times & \times & 0 & \times & \times \\ \times & \times & \times & \times & 0 & \times \\ \times & \times & \times & \times & \times & 0 \end{bmatrix}$$

Compute largest eigenvalue

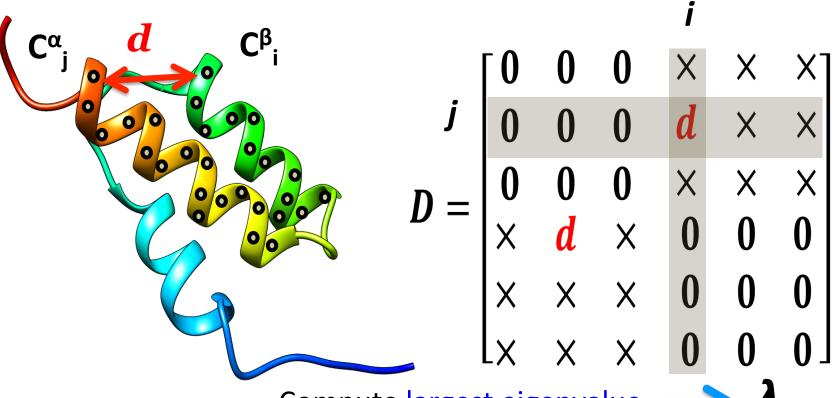




#### **Capturing Tertiary Structures**

Measure the distance between  $C^{\alpha}_{j}$  and  $C^{\beta}_{i}$ 

Build a **bipartite distance matrix** by comparing two substructures



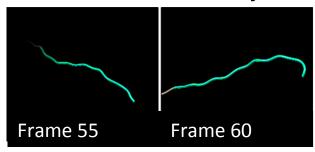
Compute largest eigenvalue

Frames of an MD job:



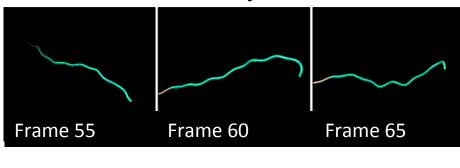
 $\lambda_{55}$ 

#### Frames of an MD job:



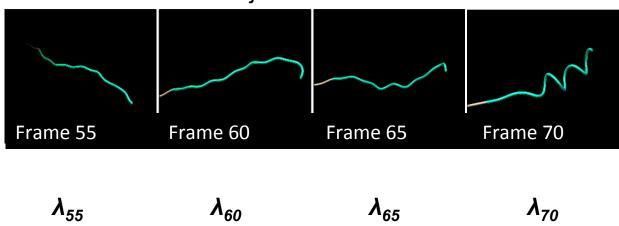
 $\lambda_{55}$   $\lambda_{60}$ 

Frames of an MD job:

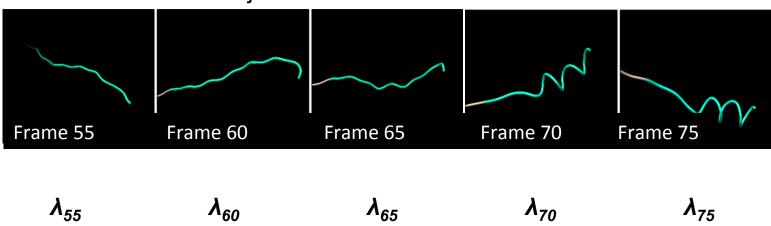


 $\lambda_{55}$   $\lambda_{60}$   $\lambda_{65}$ 

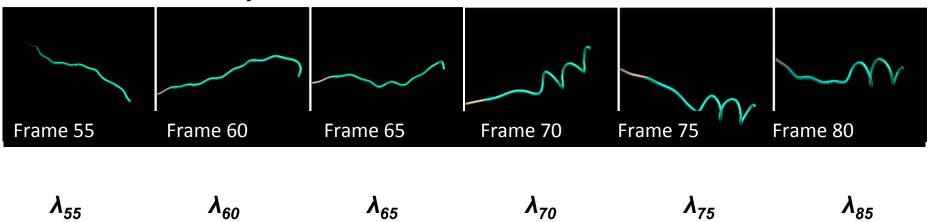
Frames of an MD job:



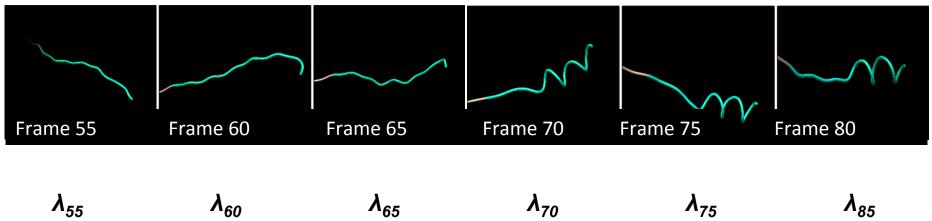
Frames of an MD job:



Frames of an MD job:



Frames of an MD job:



Distance between two max eigenvalues serves as a proxy for distance between the two associated conformations

# Distance between two max eigenvalues serves as a proxy for distance between the two associated conformations

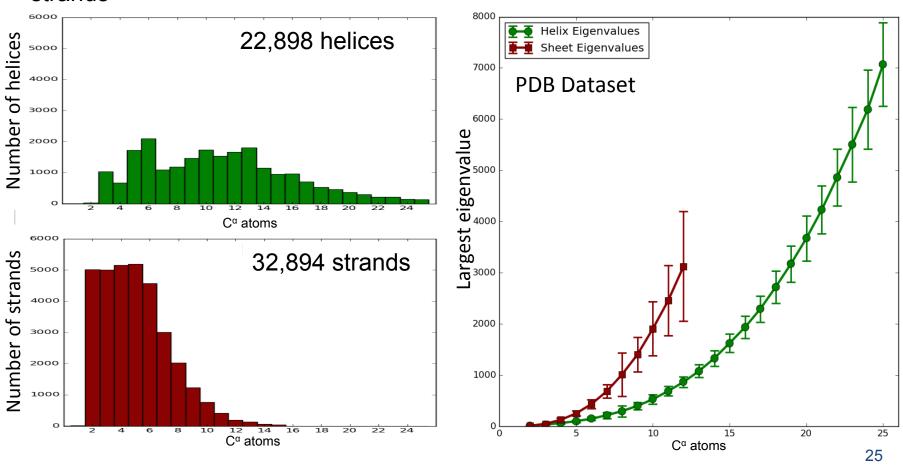
- Euclidean distance matrix D is symmetric
- Eigenvalues of symmetric, real matrices are stable
  - Small perturbations of D result in only small changes in the eigenvalues
  - Euclidean distance matrix is insensitive to rigid transformation
- Use only largest eigenvalue in distance matrix

$$\lambda max = \lambda 1 < \lambda 2 < \lambda 3 < \lambda 4 < \lambda 5 = \lambda min$$
  
 $\lambda 1 + \lambda 2 + \lambda 3 + \lambda 4 + \lambda 5 = 0$   
 $\lambda 1 >> \lambda 2 \sim \lambda 3 \sim \lambda 4 \sim 0$   
 $\lambda max = \lambda 1 \sim -\lambda 5 = -\lambda min$ 

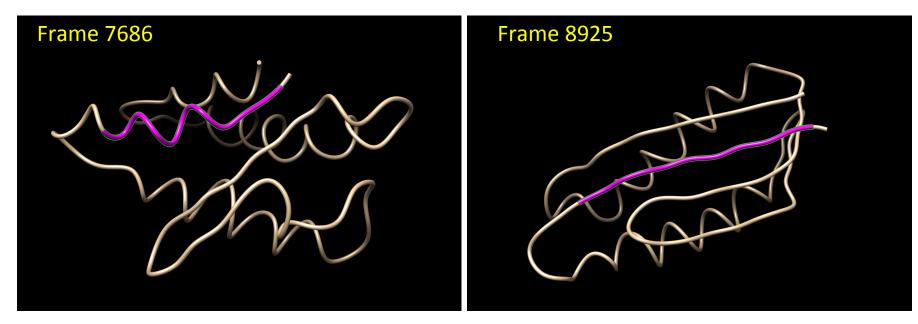
"In-Situ Data Analysis and Indexing of Protein Trajectories," Travis Johnston, Buyu Zhang, Adam Liwo, Silvia Crivelli, and Michela Taufer. JCC 2017.

#### Mapping Largest Eigenvalues to Structures

**PDB dataset:** 3,197 different proteins including 22,898 helices and 32,894 strands

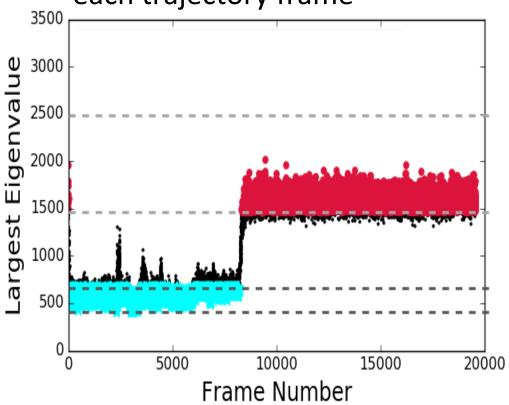


- Canonical simulation of 2MQ8 protein including both  $\alpha$  helices and  $\beta$  strands
  - After  $\sim$ 9M steps  $\alpha$  helices pack tighter and change into  $\beta$  strands

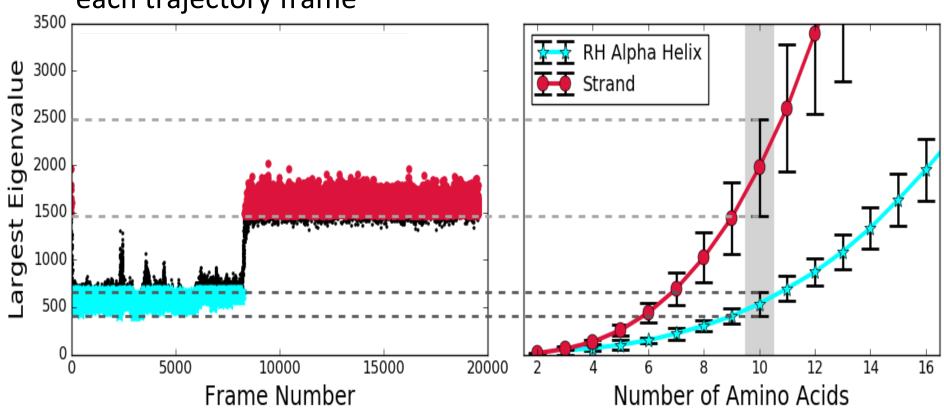


Can the eigenvalue analysis capture the conformational change?

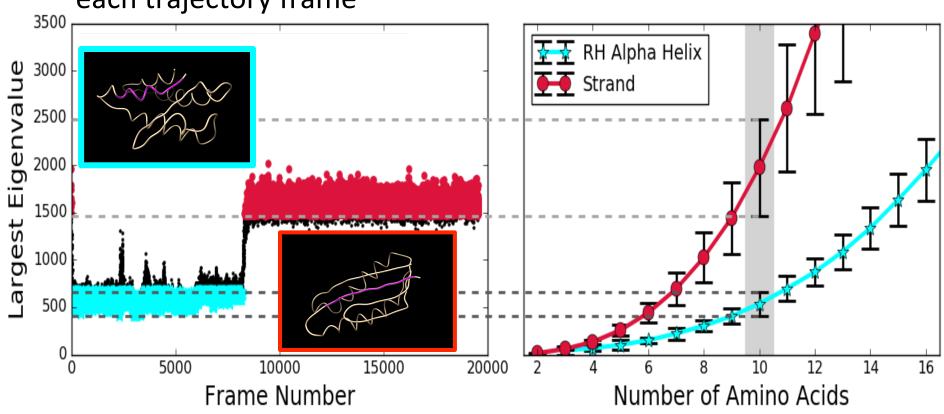
Compute largest eigenvalue of 3<sup>rd</sup> strand (10 amino acids) for each trajectory frame



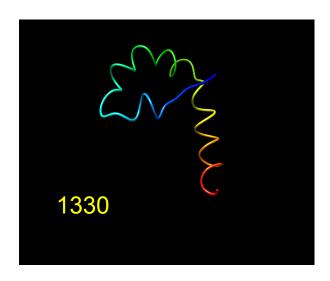
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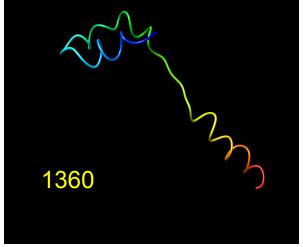


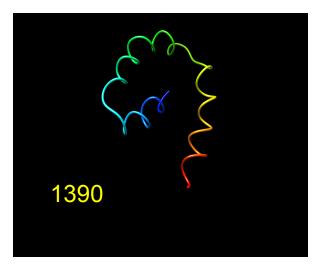
Compute largest eigenvalue of 3<sup>rd</sup> strand (10 amino acids) for each trajectory frame



Capture movement of structures with respect to each other

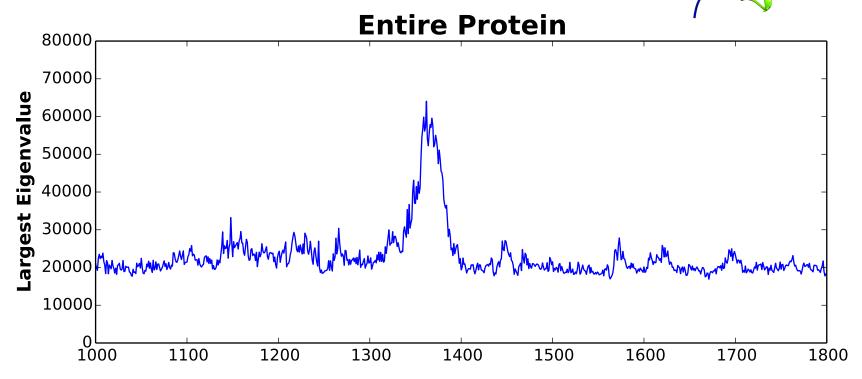




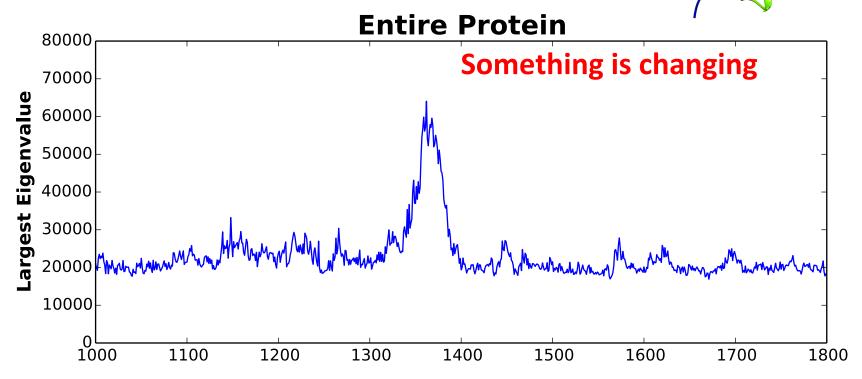


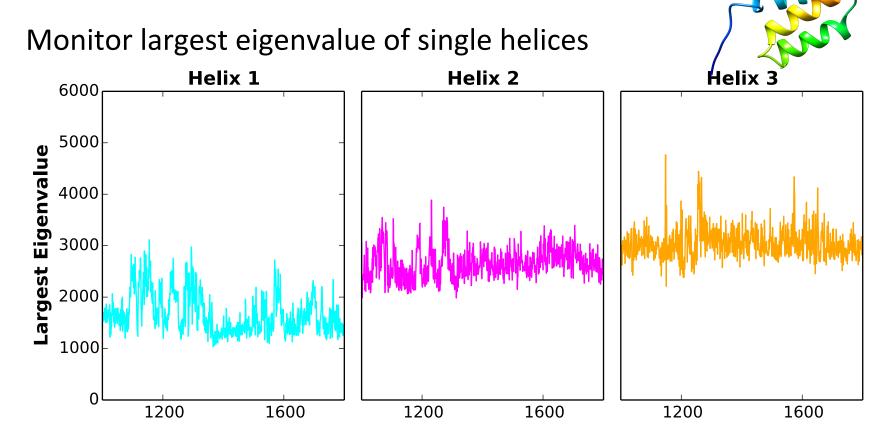
Can the eigenvalue analysis capture the movement of helices?

Monitor largest eigenvalue of entire protein



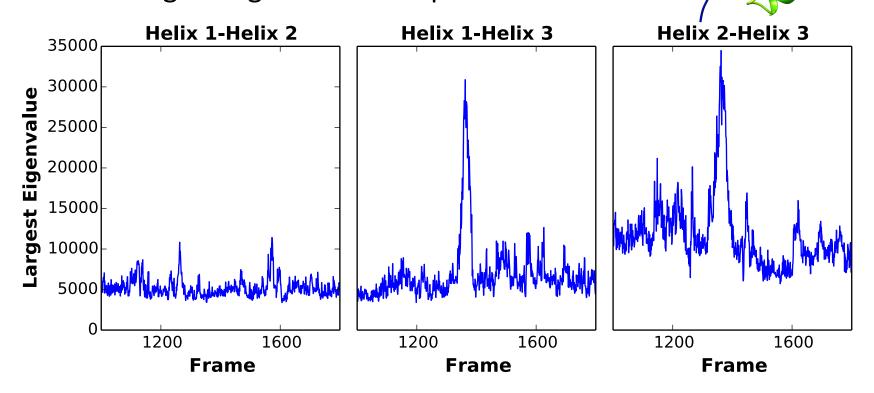
Monitor largest eigenvalue of entire protein





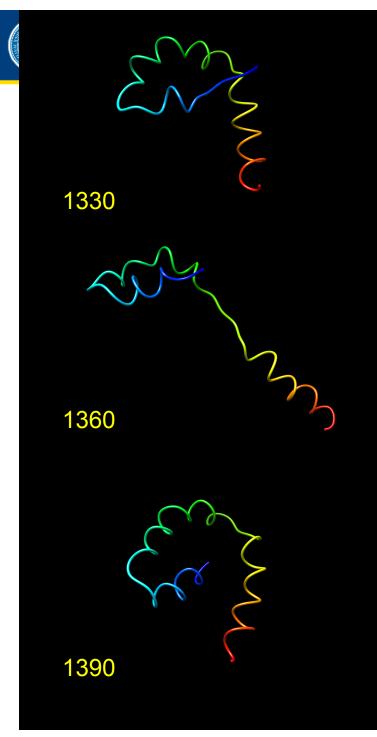
Individual  $\alpha$ -helices (Helix 1, Helix 2, and Helix 3) appear stable

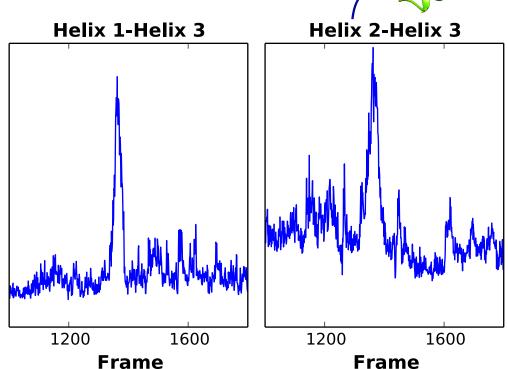
Monitor largest eigenvalue of bipartite distance matrix



First and second  $\alpha$ -helices appear stable; third helix moves







Large relative change between two pairs of α-helices

#### Perspectives

The traditional HPC point of view:

"[...] the results would not be of interest to the Supercomputing community given that the method has not been parallelized, does not discuss how the method might be parallelized, etcetera. Hence, the work may be more appropriate for a computational chemistry journal or the IEEE Big Data Conference."

Anonymous Feedback

# 10<sup>th</sup> IEEE International Scalable Computing Challenge **SCALE 2017**

- Co-located with IEEE/ACM CCGrid, Madrid, Spain, May 14-17, 2017
- Objectives:
  - Showcase real-world problems solved using computing that scales
  - Advances in applications development and infrastructure to enable scaling
- **Submissions:**

Format: White papers (6-page max. in IEEE format)

Deadline: **Jan. 31, 2017** 

All selected contributions in conf. proceedings

Awards:

First Place: \$1,000

Second Place: \$500



Algier

Melilla (Spa Sidi Bel Abbès